Prevalence of *Staphylococci* in Commercially Processed Food Products in Karachi-Pakistan

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**ABSTRACT**

**Objectives:** This study was designed to determine the prevalence of *Staphylococci*, *Staphylococcus aureus* (*S. aureus*), and Methicillin Resistant *S. aureus* (MRSA) in commercially prepared food items that involve human handling, in Karachi-Pakistan.

**Methods:** In a cross-sectional survey approach, in total 1012 food samples were analyzed. These samples were provided by different food processing industries during 2013 to 2016. Barid-Parker agar plates with egg yolk tellurite (Oxoid) along with Mannitol Salt Agar (BioM), Staph-chromo agar (Merck), Staphylococcus 110 Agar (BioM), and Blood Agar (Oxoid) were used for isolation and identification of *Staphylococci*. Polymerase chain reaction (PCR) was used for amplification of *mecA* gene and specie identification via 16s RNA.

**Results:** Total 723 samples (71.4%) showed the presence of *Staphylococci*. Out of 723 staphylococcal isolates, 367 (36.2%) were *S. aureus* and 85 (8.3%) isolates were confirmed as MRSA. Molecular studies for MRSA typing showed that most of the isolates (74.1%) belong to SCCmecA IV and 20% MRSA isolates were SCCmecA type II and 5.8% isolates carried SCCmecA type III. The MRSA isolates of SCCmecA type II belong to agr type I, while majority (67%) of the isolates carry agr type II.

**Conclusion:** This study suggested that majority of MRSA isolates recovered from commercial food items belongs to SCCmecA IV and human handling is a major factor for introduction of *staphylococci* in processed food items. *J Microbiol Infect Dis* 2017; 7(2): 83-87

**Keywords:** *Staphylococci*, *S. aureus*, Methicillin Resistant *S. aureus*

**INTRODUCTION**

*Staphylococci* are gram positive bacteria, known to survive under a wide range of environments e.g. on dry surfaces, high salt concentration and hospital set-ups [1,2]. *Staphylococci* exist in air, dust, sewage, water, milk, and food or on food equipment, environmental surfaces, humans, and animals. Humans and animals are the primary reservoirs [1]. Mostly, it colonizes persons in hospital as well as community set-ups and is transmitted via person to person contact as well as through inanimate objects [2]. The food borne cases of *Staphylococci* have been reported from all over the world [2]. Presence of pathogens in food products imposes potential hazard for consumers and causes grave economic loss via food-borne disease [3]. *Staphylococci* enter in a food set-up and contaminate the food products via human handlers or healthy nasal carriers. After entry, it multiplies in food and produce toxins that can cause food poisoning, a gastrointestinal illness [2,3]. Food items made by hands without cooking are at high risk to carry *Staphylococcal* toxins. However air, dust, and food contact surfaces may also serve as vehicles in the transfer of *Staphylococci* in foods. *Staphylococcus aureus* is one of the well-known pathogen of *Staphylococci* family [1-3]. It tolerates high salt concentration; desiccation and can grow in a wide range of temperatures (7 °C to 48.5 °C; optimum 30 to 37 °C), pH (4.2 to 9.3; optimum 7 to 7.5). These characteristics favor growth of the organism in many food products [3]. The highly resistant and pathogenic
strain among Staphylococci is Methicillin Resistant S. aureus (MRSA). Originally, MRSA was found in hospital set-ups; in recent times it is frequently reported in the community as well, and considered as a source of food borne illness [4]. Food borne MRSA isolates are resistant to penicillin and oxacillin, only, whereas hospital isolates of MRSA are multi-drug resistant hence are difficult to control [4]. In S. aureus Methicillin resistance is mediated by mecA gene, which is located on mobile genetic element known as staphylococcal cassette chromosome mec (SCCmec). There are five major types of SCCmec elements (I-V). The majority of hospital-acquired MRSA strains carry SCCmec types I, II, or III, whereas community-acquired MRSA strains carry SCCmec types IV or V [8,9]. World Health Organization (WHO) defines food-borne disease as “disease of infectious or toxic nature caused by the consumption of food or water” [3]. S. aureus is one of the major pathogens responsible for food borne infections, world-wide. Present study describes the prevalence of Staphylococci, S. aureus and MRSA in different food items that involve human handling.

METHODS
In present study total 1012 food samples were analyzed. These samples were provided by different food processing industries during 2013 to 2016.

Isolation and Enumeration of Staphylococci and S. aureus
Fifty grams of each sample were mixed with 450 ml of 0.1% peptone sterile physiological saline solution (0.85% NaCl) and homogenized. Three decimal dilutions of each sample homogenate were prepared for enumeration of S. aureus. From each dilution, 0.3, 0.3 and 0.4 ml was spread on Barid-Parker agar plates with egg yolk tellurite and were kept in an upright position until liquid was absorbed and then incubated at 35 °C for 24–48 h. Typical black colonies with white zone were considered as S. aureus [5]. Staph Latex Kit (Prolix Latex Agglutination System) and growth on Mannitol Salt Agar (BioM), Staph-chromo agar (Merck), Staphylococcus 110 Agar (BioM), and Blood Agar (Oxoid) was used for further confirmation.

Determination of plasma-coagulase and nuclease activity
Colonies of staphylococci were individually re-inoculated into test tubes containing 1.5 mL of Brain Heart Infusion Broth (BHI Broth, Oxoid). After 24h of incubation at 35°C, 0.5 mL of each sample was added to another test tube containing 1 mL rabbit plasma (Merck). Inoculated test tubes were incubated at 35°C. Formation of coagulum was considered as positive reaction. Results were evaluated after 1, 2, 3, 6 and 24 h. Each isolate was inoculated on the surface of DNase Agar (Oxoid) and was incubated at 35°C. After 24h of incubation, medium was flooded and acidified with 1 N hydrochloric acid, the DNA precipitated the turbidity of medium and clear zones around colonies indicated positive DNase reaction. The number of isolates that presented positive and negative results was recorded [10].

Determination of MIC for oxacillin
BHI Agar (Oxoid) was used to measure the oxacillin resistance level according to the guidelines of Clinical Laboratory Standard Institute (CLSI) [6]. Minimum inhibitory concentration (MIC) was re-confirmed by E-test using AB-Biodisk according to the manufacturer's instructions.

Polymerase chain reaction (PCR)
For molecular studies, genomic DNA was isolated by using the DNase Kit (Qiagen), following the manufacturer's instructions. PCR amplification of mecA genes was performed with an MWG Thermal Cycler in a volume of 50 μl of Promega Master Mix. Primers, described previously [6], were used for amplification of mecA, for SCCmec typing of MRSA isolates and for agr allele (I-IV) typing. 16S RNA was used as internal control for gene expression and specie identification.

Ethical approval
This study was approved by the institutional ethics committee.

RESULTS
Present study describes the presence of Staphylococci in different food products. Specially, foods that require considerable handling during preparation and that are kept at slightly elevated temperatures after preparation...
are selected and included in this study (Table 1). Total 1012 samples were analyzed, 723 (71.4%) samples showed the presence of Staphylococci (Table 1, Figure 1).

Out of 723 staphylococcal isolates, 367 (36.2%) were S. aureus and 85 (8.3%) isolates were confirmed as MRSA (Table 1, Figure 1). These isolates were identified on the basis of growth characters on selective and differential media. An isolate was considered within Staphylococcus genus when presented as gram-positive cocci in grape-like clusters, catalase, and mannitol fermentation positive. Coagulase test and growth on Baird-Parker agar and DNase agar confirmed the presence of S. aureus in subject food items. Positive reaction or amplification of 16s and mecA gene specific primers confirm the presence of MRSA isolates in 85 food items. Out of 1012 samples 338 were spices mix; 80.77% of these samples showed the presence of staphylococci. The pathogenic S. aureus were present in 43.79% of spices mix samples with 5.03% of MRSA isolates. Among candy samples, 80.95% were positive for Staphylococci; 23.81% of these were S. aureus and 9.52% were MRSA. In fish and meat products, 40% and 87% showed Staphylococcal growth, respectively. In fish products S. aureus were 28% with 16% MRSA and in meat products 39% S. aureus with 7% MRSA. Samosa and Paratha (Frozen products) showed Staphylococci in 55% and 64% of samples, respectively. In Parathas 30.67% isolates were identified as S. aureus, in Samosa the S. aureus were 45%, with 18.67% and 10%, respectively MRSA. Rice and lentils respectively, showed 40.37% and 27.5% S. aureus with 9.17% and 7.5% MRSA. About 20% samples of Formula Milk and 10% of Mayonnaise samples were positive for MRSA. Table 1 depicts the complete details of samples and isolates. Moreover, molecular studies for MRSA typing showed that most of the isolates (74.1%) belong to SCCmecA IV and 20% MRSA isolates were SCCmecA type II and 5.8% isolates carry SCCmecA type III. Interestingly, isolates recovered from formula milk and mayonnaises were SCCmecA type II. The isolates that carry SCCmecA type III were recovered from meat products and candy samples. The MRSA isolates of SCCmecA type II belong to agr type I, while majority (67%) of the isolates carry agr type II.

### Table 1. MICs and typing of SCCmecA genes and agr in MRSA isolated from different food products.

<table>
<thead>
<tr>
<th>Source</th>
<th>No. of Samples</th>
<th>Staphylococci, n (%)</th>
<th>S. aureus, n (%)</th>
<th>MRSA, n (%)</th>
<th>MICs (µ/ml)</th>
<th>AGR Typing</th>
<th>SCCmecA Typing</th>
</tr>
</thead>
<tbody>
<tr>
<td>Candies</td>
<td>105</td>
<td>85 (80.95)</td>
<td>25 (23.81)</td>
<td>10 (9.52)</td>
<td>2 4 4 2 8 2</td>
<td>2 2 6</td>
<td></td>
</tr>
<tr>
<td>Candy mix</td>
<td>50</td>
<td>40 (80.00)</td>
<td>19 (38.00)</td>
<td>4 (8.00)</td>
<td>0 4 0 0 4 0</td>
<td>0 0 0 4</td>
<td></td>
</tr>
<tr>
<td>Dates</td>
<td>85</td>
<td>40 (56.25)</td>
<td>17 (20.00)</td>
<td>6 (7.06)</td>
<td>2 4 0 0 6 0</td>
<td>0 0 0 6</td>
<td></td>
</tr>
<tr>
<td>Fish Products</td>
<td>25</td>
<td>10 (40.00)</td>
<td>07 (28.00)</td>
<td>4 (16.00)</td>
<td>0 0 4 0 4 0</td>
<td>0 0 0 4</td>
<td></td>
</tr>
<tr>
<td>Formula Milk</td>
<td>15</td>
<td>10 (66.67)</td>
<td>08 (53.33)</td>
<td>3 (20.00)</td>
<td>0 0 3 3 0 3</td>
<td>0 0 0 0</td>
<td></td>
</tr>
<tr>
<td>Lentils</td>
<td>40</td>
<td>19 (47.50)</td>
<td>11 (27.50)</td>
<td>3 (7.50)</td>
<td>2 1 0 1 2 0</td>
<td>0 0 3</td>
<td></td>
</tr>
<tr>
<td>Mayonnaise</td>
<td>30</td>
<td>12 (40.00)</td>
<td>08 (26.67)</td>
<td>3 (10.00)</td>
<td>0 3 0 3 0 3</td>
<td>0 0 0</td>
<td></td>
</tr>
<tr>
<td>Meat Products</td>
<td>100</td>
<td>87 (87.00)</td>
<td>39 (44.83)</td>
<td>7 (7.00)</td>
<td>2 5 0 3 4 0</td>
<td>3 4 3 4</td>
<td></td>
</tr>
<tr>
<td>Paratha</td>
<td>75</td>
<td>48 (64.00)</td>
<td>23 (30.67)</td>
<td>14 (18.67)</td>
<td>4 7 3 6 8 3</td>
<td>0 0 11</td>
<td></td>
</tr>
<tr>
<td>Rice</td>
<td>109</td>
<td>77 (70.64)</td>
<td>44 (40.37)</td>
<td>10 (9.17)</td>
<td>0 5 5 2 8 0</td>
<td>0 0 10</td>
<td></td>
</tr>
<tr>
<td>Samosa</td>
<td>40</td>
<td>22 (55.00)</td>
<td>18 (45.00)</td>
<td>4 (10.00)</td>
<td>4 0 0 4 0 2</td>
<td>0 0 2</td>
<td></td>
</tr>
<tr>
<td>Spices Mix</td>
<td>338</td>
<td>273 (80.77)</td>
<td>148 (43.79)</td>
<td>17 (5.03)</td>
<td>17 0 0 0 17 0</td>
<td>0 0 17</td>
<td></td>
</tr>
</tbody>
</table>

### DISCUSSION

In the present study, commercial food items, those involve human handling, during processing and packaging, showed high rate of staphylococcal contamination, including S. aureus and MRSA. In present study it has been observed that food handlers are the major...
source of staphylococcal contamination. Although, the original source for staphylococcal introduction to food items was not traced out, even then this study creates doubt about food safety and highlights influence of food handling and processing on consumer health. Of more concern is the presence of MRSA; out of 1012 samples 367, (36.2%) were contaminated with S. aureus and 85 (8.3%) with MRSA. This is a very serious situation. Due to multi-drug resistance and enterotoxins production, MRSA could be more fatal as compared to methicillin sensitive S. aureus. According to Jones at al [7] MRSA are as likely to produce enterotoxins as are methicillin-sensitive strains. Normally, it is considered that MRSA can survive in limited food items e.g. dairy products, meat and through cross contamination during kitchen processing of the recipe. However, during present study, recovery of MRSA from different variety of foods e.g. Lentils, Rice, Spices mix, Sweet mix and Dates, suggest the versatility of this pathogen. Moreover, majority of these isolates exhibited low-level oxacillin resistance (MIC ranges 32 to 128µg/ml), a character of the community of MRSA. According to Boyle-Vavra et al. [8], community-acquired MRSA (CAMRSA) isolates usually carry SCCmec type IV. Out of 87 subject isolates of MRSA, 63 (72.4%) belong to SCCmec type IV. According to Song et al. [9] SCCmec type IV is the most predominant community type clone in the Asian countries. Interestingly, MRSA that belongs to SCCmec type II was recovered from sweet products only e.g. candies, mayonnaise, and formula Milk; these products involve more human handling as compared to the other products tested. The SCCmec type IV normally belongs to community acquired isolates of MRSA and SCCmec type II is associated with hospital acquired isolates. It is reported that SCCmec type II isolates are highly resistant type of MRSA, whereas SCCmec type IV exhibited low-level of resistance and are only resistant to β-lactam antibiotics; as noticed in the present study. Although, this study is based on food items that involve human handling, but the true source of MRSA remains to be elucidated. Of more concern is to determine how these isolates of S. aureus develop oxacillin resistance. According to best of our knowledge, in Pakistan the use of antibiotics in agriculture is not common. However, use of antibiotics as a growth promoter in poultry industry is a common practice but good data are not available. So, this study which is based on wide range of samples provides us intimation about the prevalence rate of MRSA is our community.

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Competing interests:

None to declare.

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